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Provisional specification in connection with Application No. PS 1118 for a
patent by PROTEOME SYSTEMS INTELLECTUAL PROPERTY PTY LTD as
filed on 13 March 2002.



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Mathew Danger Traini

Title: ANNOTATION OF GENOME SEQUENCES

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PROVISIONAL SPECIFICATION

Invention Title:

Annotation of genome sequences

The invention is described in the following statement:

Field of the Invention

This invention relates to a method of annotation of genome sequences.

Background of the Invention

5 Many genomes, including the human genome have now been sequenced. A genome sequence provides a list of bases (A,T,G,C) in the order in which they appear in a length of DNA, however, the sequence *per se* tells one very little about the genome that is useful and easily or immediately comprehensible. For example in the study of a disease causing bacteria it
10 would be useful in searching for a cure for the disease to determine the location of that part of the bacterium's genome which expressed a particular protein. However, it can be difficult to predict where proteins of interest may be located in a genome sequence. It cannot always be done simply by looking at the sequence *per se*.

15 There are a number of known processes for attempting to determine the location of proteins in genome sequence data. One known method uses computer programs to locate recognisable regions such as start codons and stop codons in a DNA sequence. Other programs attempt to locate proteins by locating regions of high complexity within a DNA sequence which typically
20 indicates the location of a protein.

However, these approaches are far from perfect as in order to implement these programs, various assumptions and hypotheses have to be made about the location of a protein of interest in the DNA sequence, in particular, the potential start and stop positions of the protein. A detection method that
25 requires such assumptions or hypotheses may produce incorrect results if the assumptions/hypotheses are incorrect. For example these procedures are unlikely to locate non-typical sequences, which ironically may be of more interest than other proteins having more typical sequences identified using existing techniques.

30 Thus, it is one object of the present invention to provide a method for annotating genome sequences, which is hypothesis independent and does not make assumptions for the detection of a protein from nucleic acid sequences.

Any discussion of documents, acts, materials, devices, articles or the like which has been included in the present specification is solely for the purpose of
35 providing a context for the present invention. It is not to be taken as an admission that any or all of these matters form part of the prior art base or were

common general knowledge in the field relevant to the present invention as it existed in Australia before the priority date of each claim of this application.

Summary of the Invention

5 A first broad aspect of the present invention, provides a method of identifying one or more proteins in an unannotated DNA sequence, the method comprising:

(a) dividing the DNA sequence into a plurality of sequence fragments each fragment being of substantially the same length and from about 300 to
10 5000 base pairs long;

(b) performing a six frame translation of each of the DNA sequence fragments to obtain six translated amino acid sequence fragments for each DNA sequence fragment;

(c) subjecting each of the translated sequence fragments to theoretical
15 digestion to obtain a plurality of cleaved peptide sequences;

(d) comparing experimental empirical data for peptide fragments from a protein digested in the same manner as the theoretical digestion at step (c) with the theoretical data generated in step (c) for each of the translated sequence fragments to identify one or more translated sequence fragments
20 which include a substantial number of peptides present in the digested protein.

An advantage of the present invention is that no assumptions need to be made about the location of proteins in the DNA sequence data. DNA sequences with non-typical stop and or start codons may be located. The results are hypothesis independent.

25 Typically the theoretically generated peptide masses are compared to the masses of the peptides experimentally generated by the digested protein and the sequence fragment which has the greatest number of theoretical peptide masses correlating to the empirical data indicates the likely location of the protein of interest in the DNA sequence. The masses of the peptides
30 experimentally generated from the digested protein will typically be determined by mass spectrometry.

It is preferred that the DNA sequence is duplicated and the original and duplicate are split in such a manner that the sequence fragments from the original overlap the cuts in the original genome sequence.

35 It is important that the sequence fragments are approximately the same length as one another and are sized to equate to the length of a typical protein.

Hence, each fragment is, as discussed above, about 300-5000 base pairs long. Proteins vary in size, most proteins being 10 to 100 kDa i.e. about 300-3000 base pairs long. Most preferably, the sequence fragments will be around 1000 or 1050 bases long, the latter translating to 350 amino acids which is approximately equivalent to a 33 to 37 kDa protein, which is a common size for a protein.

Using DNA sequences of approximately that length produce about 12 to 20 hits against a background number of hits of around 4 for sequences which do not contain a protein.

10 In a related aspect of the present invention, the step of dividing the DNA sequence and the step of performing the six frame translation can be reversed. Hence, a second broad aspect of the present invention provides a method of identifying one or more proteins in unannotated DNA sequence, the method comprising:

15 (a) performing a six frame translation of a DNA sequence to provide six translated amino acid sequences;

(b) dividing the six translated amino acid sequences into a plurality of fragments, each fragment comprising 100-1666 amino acids;

20 (c) subjecting each of the fragments to theoretical digestion to obtain a plurality of cleaved peptide sequences;

(d) comparing experimental empirical data for peptide fragment for peptide fragments from a protein digested in the same manner as the theoretical digestion at step (c) with theoretical data generated in step (c) for each of the fragments to identify one or more fragments which include a substantial number of peptides present in the empirically digested protein.

Brief Description of the Drawings

A specific embodiment of the present invention will now be described by way of example with reference to the accompanying drawings in which:

30 Figures 1A to 1E are schematic diagrams illustrating various steps in the method of the present invention;

Figure 2 shows a report comparing empirical masses of the protein Glycerol-3-phosphate dehydrogenase (P95113) against a split, translated, and digested genome sequence of M Tuberculosis;

Figure 3 shows a detailed report for the frame which provided the greatest number of hits (eighteen) comparing the theoretical masses of each peptide in the digest with the empirical masses; and

Figure 4 shows detailed reports for two frames which have provided only
5 four hits;

Figure 5 shows detailed results for one frame/fragment number 6366; and

Figure 6 shows detailed results for another frame/fragment number 6364.

10

Detailed Description of a Preferred Embodiment

Referring to the drawings, Figure 1A, shows a genome sequence 10 which is taken and split into a series of shorter genome sequences or sequence fragments 12. Overlapping sequences are preferably provided by
15 duplicating the genome sequence and cleaving the duplicated sequence at locations midway between the breaks in the original sequence so that the sequences (12a,12b..., 14a, 14b...) are overlapping as shown in Figure 1A.

Typically, the genome will be cut into sequence fragments which are 1050 bases long. This approximates to 350 amino acids which will be found in
20 a protein of around 33 to 37 kDa which is a common protein size. A bacterium such as Mycobacterium tuberculosis (Tb) will have around 4.4 million bases in its genome. Duplicating and cutting that genome will result in approximately 8400 sequence fragments.

A six frame translation is then carried out on each of the sequence
25 fragments. Figure 1B schematically illustrates a 6 frame translation carried out on one of the sequence fragments (14d). For each fragment, six virtual proteins are produced. Fragment 14d produces six virtual proteins 16a-16g. Using the M Tuberculosis example referred to above the 8400 virtual proteins become 50,400. These virtual proteins are then subjected to theoretical
30 digestion according to rules which mimic the action of an endoproteinase enzyme such as trypsin which cut at specific target sites on a target sequence. This digestion is schematically illustrated in Figure 1C. Each virtual protein becomes a series of "virtual peptides" and the mass of each virtual peptide is calculated. "Protein" 16g becomes six peptides 18a to 18g. Fewer or more
35 peptides may be produced from each virtual protein. The protein of interest is then subjected to an empirical digestion using the same enzyme and peptide

mass data is obtained from mass spectrometry of the peptides expressed by that protein.

The masses of the various empirically derived peptides are then compared with the theoretical peptide masses produced by theoretical
5 cleavage of the sequence fragments. This is done in a stepwise manner and frame by frame whereby all the empirical peptide masses are matched against all peptides from the first virtual protein and the number of matching peptides (hits) is recorded. For each virtual protein, this process is carried out six times, once for each of the amino acid translations. However, the number of hits for
10 each frame is calculated separately and the hits are not summed together. This process is then repeated for the second virtual protein and so on, until it has been carried out for all the virtual proteins. This step is illustrated in Figure 1D. There is a background number of hits. Typically, each theoretical protein or sequence fragment will produce about 3 or 4 peptides having masses which
15 correlate to masses produced by the actual empirical digest of the protein of interest. The sequence fragment which produced the protein of interest will in contrast typically have 12 to 20 peptide matches with the empirical digest of the protein of interest but is limited by the number of peptides generated empirically.

20 Clearly the relevant part of the genome sequence may have been cut in the original division of the genome sequence, however the overlapping of the original and duplicate genome sequences reduces the risk of this. In any case even if the protein is split it is still possible to identify the relevant part of the genome sequence because we would see a reasonable number e.g. 6 to 10
25 hits in two adjacent overlapping fragments. The part of the sequence which carries the most peptide masses which match the peptide masses produced by the empirical digestion and has a number of hits which is clearly above the background (noise) level is likely to be that part of the genome which carries the protein of interest. By knowing where the part of the sequence came from,
30 this identifies the location of the protein in the genome sequence (Figure 1E).

The present invention works particularly well with small genomes such as bacterial and yeast genomes or other eukaryote genomes that have few introns and small amounts of non-coding DNA.

The method can also be used for the detection of pseudo genes which
35 are versions of genes which have become defunct and identifying "protein families" of similar proteins. When a protein from a family of proteins is

detected, a number of regions having a large number of matches may be identified. This indicates that the proteins may be members of the same protein family which may be for example be expressed in different tissues. The method also identifies if there is a now inactive or defunct family region of genome to a related region that carries an active version of a protein.

Example

Figures 2 to 6 illustrate the results of carrying out the method of the present invention looking for the part of the M Tuberculosis genome which produces the protein Glycerol-3-phosphate dehydrogenase (P95113). The protein was digested with trypsin and the masses of the peptides produced by the digestion measured using mass spectrometric analysis.

The empirical masses were searched against the split, translated and theoretically digested genome sequence for M Tuberculosis using the method of the present invention as discussed above. Figure 2 shows a summary of the results illustrating all the theoretical sequence fragments which produced four hits or more. One fragment 6365 derived from the genome base pairs 3341051 to 3342100 produced 17 hits. This fragment contains the complete sequence for the protein of interest. Two adjacent fragments 6364 and 6366 produced eight hits each, as they are overlapping and contain part of the protein of interest. Other results show a background number of 4 or 5 hits only.

Figure 3 is a detailed report on segment 6359. For each peptide in the digest, the theoretical database mass is compared with the empirical mass determined by mass spectrometry. The start and end positions of each of the peptides in the sequence are also given. Note that in Figure 2, the peptides are ordered by increasing mass. They could simply be ordered by position in the sequence. The report also indicates if any cleavages have been missed (MC) and the frame number "5" indicates that the fifth frame (out of the six frame translations of the genome fragment of interest) produced the match with the empirical data.

The sequence is shown at 50 at the top of the report. Note that there are no stop codons 52 marked with an asterisk in the areas where the protein is located.

In contrast, Figure 4 shows the reports on two of the fragments (numbers 238 and 749) which had four hits only - a background number of hits. The matching peptides are scattered throughout the sequence. In the case of

fragment 238 there are frequent stop codons suggesting that this is not a coding region of the genome. For fragment 749 there is a background number of hits to what is probably a coding region of the genome, but coding a protein different to the one we are analysing.

5 Figure 5 shows detailed results for fragment number 6366 which had 8 hits. This shows that a portion of the protein sequence is present in the second half of this fragment. Figure 6 shows detailed results for fragment number 6364. this shows that a portion of the protein sequence is present in the first half of this fragment.

10 It will be appreciated by persons skilled in the art that numerous variations and/or modifications may be made to the invention as shown in the specific embodiments without departing from the spirit or scope of the invention as broadly described. The present embodiments are, therefore, to be considered in all respects as illustrative and not restrictive.

Dated this thirteenth day of March 2002

Proteome Systems Intellectual
Property Pty Ltd
Patent Attorneys for the Applicant:

F B RICE & CO

genomic sequence

split into overlapping fragments

Figure 1A

translate in six frames

Figure 1B

Figure 1C

theoretical enzymatic digestion

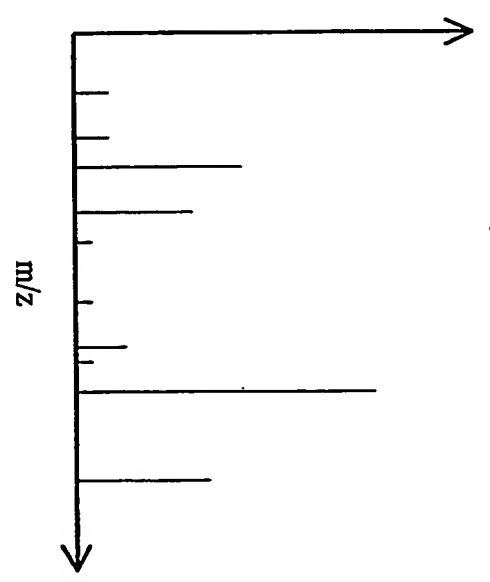
comparison with experimental data

Figure 1D

location of protein in genome sequence

Figure 1E

experimental enzymatic digest



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Prepared: B. B. Gagnon, Assistant to Director, Bureau of Census
 and Statistics, Department of Commerce
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Accession number	Protein name	Q1	Q2	Q3	Q4	Q5	Q6	Q7	Q8	Q9	Q10	Q11	Q12	Q13	Q14	Q15	Q16	Q17	Q18	Q19	Q20	Q21	Q22	Q23	Q24	Q25	Q26	Q27	Q28	Q29	Q30	Q31	Q32	Q33	Q34	Q35	Q36	Q37	Q38	Q39	Q40	Q41	Q42	Q43	Q44	Q45	Q46	Q47	Q48	Q49	Q50	Q51	Q52	Q53	Q54	Q55	Q56	Q57	Q58	Q59	Q60	Q61	Q62	Q63	Q64	Q65	Q66	Q67	Q68	Q69	Q70	Q71	Q72	Q73	Q74	Q75	Q76	Q77	Q78	Q79	Q80	Q81	Q82	Q83	Q84	Q85	Q86	Q87	Q88	Q89	Q90	Q91	Q92	Q93	Q94	Q95	Q96	Q97	Q98	Q99	Q100	Q101	Q102	Q103	Q104	Q105	Q106	Q107	Q108	Q109	Q110	Q111	Q112	Q113	Q114	Q115	Q116	Q117	Q118	Q119	Q120	Q121	Q122	Q123	Q124	Q125	Q126	Q127	Q128	Q129	Q130	Q131	Q132	Q133	Q134	Q135	Q136	Q137	Q138	Q139	Q140	Q141	Q142	Q143	Q144	Q145	Q146	Q147	Q148	Q149	Q150	Q151	Q152	Q153	Q154	Q155	Q156	Q157	Q158	Q159	Q160	Q161	Q162	Q163	Q164	Q165	Q166	Q167	Q168	Q169	Q170	Q171	Q172	Q173	Q174	Q175	Q176	Q177	Q178	Q179	Q180	Q181	Q182	Q183	Q184	Q185	Q186	Q187	Q188	Q189	Q190	Q191	Q192	Q193	Q194	Q195	Q196	Q197	Q198	Q199	Q200	Q201	Q202	Q203	Q204	Q205	Q206	Q207	Q208	Q209	Q210	Q211	Q212	Q213	Q214	Q215	Q216	Q217	Q218	Q219	Q220	Q221	Q222	Q223	Q224	Q225	Q226	Q227	Q228	Q229	Q230	Q231	Q232	Q233	Q234	Q235	Q236	Q237	Q238	Q239	Q240	Q241	Q242	Q243	Q244	Q245	Q246	Q247	Q248	Q249	Q250	Q251	Q252	Q253	Q254	Q255	Q256	Q257	Q258	Q259	Q260	Q261	Q262	Q263	Q264	Q265	Q266	Q267	Q268	Q269	Q270	Q271	Q272	Q273	Q274	Q275	Q276	Q277	Q278	Q279	Q280	Q281	Q282	Q283	Q284	Q285	Q286	Q287	Q288	Q289	Q290	Q291	Q292	Q293	Q294	Q295	Q296	Q297	Q298	Q299	Q300	Q301	Q302	Q303	Q304	Q305	Q306	Q307	Q308	Q309	Q310	Q311	Q312	Q313	Q314	Q315	Q316	Q317	Q318	Q319	Q320	Q321	Q322	Q323	Q324	Q325	Q326	Q327	Q328	Q329	Q330	Q331	Q332	Q333	Q334	Q335	Q336	Q337	Q338	Q339	Q340	Q341	Q342	Q343	Q344	Q345	Q346	Q347	Q348	Q349	Q350	Q351	Q352	Q353	Q354	Q355	Q356	Q357	Q358	Q359	Q360	Q361	Q362	Q363	Q364	Q365	Q366	Q367	Q368	Q369	Q370	Q371	Q372	Q373	Q374	Q375	Q376	Q377	Q378	Q379	Q380	Q381	Q382	Q383	Q384	Q385	Q386	Q387	Q388	Q389	Q390	Q391	Q392	Q393	Q394	Q395	Q396	Q397	Q398	Q399	Q400	Q401	Q402	Q403	Q404	Q405	Q406	Q407	Q408	Q409	Q410	Q411	Q412	Q413	Q414	Q415	Q416	Q417	Q418	Q419	Q420	Q421	Q422	Q423	Q424	Q425	Q426	Q427	Q428	Q429	Q430	Q431	Q432	Q433	Q434	Q435	Q436	Q437	Q438	Q439	Q440	Q441	Q442	Q443	Q444	Q445	Q446	Q447	Q448	Q449	Q450	Q451	Q452	Q453	Q454	Q455	Q456	Q457	Q458	Q459	Q460	Q461	Q462	Q463	Q464
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[illegible]

98%

Local Internet

Figure 2

Sequence name: Genome-base pairs 3341051 - 3342100

Isoelectric point: 6.26 (Calton EST)

TYPE 15HCP ARPTAAR

Amino acid coverage: 317 Percentage coverage: 91.05%

১০

[illegible]

Accession number: 238 abest
Species: Molecule mass: 38340.78 Dalton EST
Sequence name: Genome base pairs 124401 - 125450
Isoelectric point: 11.21 Caudon EST

Amino acid coverage: 31 Percentage coverage: 8.21%

Database ID	Database Mass	User Mass	Difference	Start	End	WLC	Frame	Sequence
109658	60238772	60232618	0.062	321	325	0	5	QTLK
109659	65736838	65740476	-0.036	23	28	0	5	QPLSGR
109642	8904518	89045831	-0.007	85	92	0	5	NLTVGCCQ
109644	123757618	123761238	-0.036	119	130	0	5	QDAVDAGDSR

Accession number: 749 abest
Species: Molecule mass: 38514.65 Dalton EST
Sequence name: Genome base pairs 392701 - 393750
Isoelectric point: 10.24 Caudon EST

HARRSLLEP ATRRLARGL RDHSLVARE HITRORHAI INSLEHNAI TLDTILRVF GUTDPKVAE LTRSLQILN THPLILIGVF YPSLRKRNPU KRYFHNOTKI
DELIVRELS RHDSRLIAR TVUSRIQT KDPTRELD AELHDLITL LLAGETIAA ALSHLELA HAPLOSQV GAAGGDGGF LEAVLEGRH RHIVASTAR
KTLPLATGG TRLPAGTVA TSILAHASE VEHKTEER PENTLDGSA PNTSLPCCG VRKLGTFGA LLEGAVILOE ITRRTITAA GPKERTPLV RHITVPRKG
MHLNLPQNR LGGLKDSQD

Amino acid coverage: 33 Percentage coverage: 9.43%

99% Local View

Figure 4

186338	2627 3826	0 000	35	63	0	6	IGTALGANGATTACIAGVGDVATCTSPR
348864994	3488 64994	0 000	85	117	0	6	ACHVEGVTSCESTALASSIDVENMETDAVHR
186341							

1665 95791 988 0354453 20523 03 144682603 23630

Accession number: 6366 abest

Sequence name: Genome base pairs: 3341601 - 3342030

Species: *Molecular mass: 34930.68* Caution: **EST**

Isoelectric point: 8.28 **Caudon:** EST

KSPCSTTLE AP4L7TSMAL VNAVTSGSGS SSGVOSASTS APSPAAAAA ASSGVITVE RSEPTPLAAV NESTNLSSTT FSRVCAENGT ASJVLAAAR LTANIRPSP
 SSGVLEHS CORSTASTS TALPPSPBPG PRGDDGNG AB CVEDACI ASTVAVNGAG ABGTALANTL ADAGGAVTLA ARARAVADOL NITKEDBYL PALLPSPSTH
 AYADDEALNG GASVALL GVE AUMORALIER NADLLEBAT LVSLANGIEL GTLHRASQVY ISVTBAEPQ VAVISQENHA SEDACQUPHA TVVACDSOR AVALLNLSNS
 GTTRJTNAD VVGTICG

Amino acid coverage: 147 Percentage coverage: 42.24%

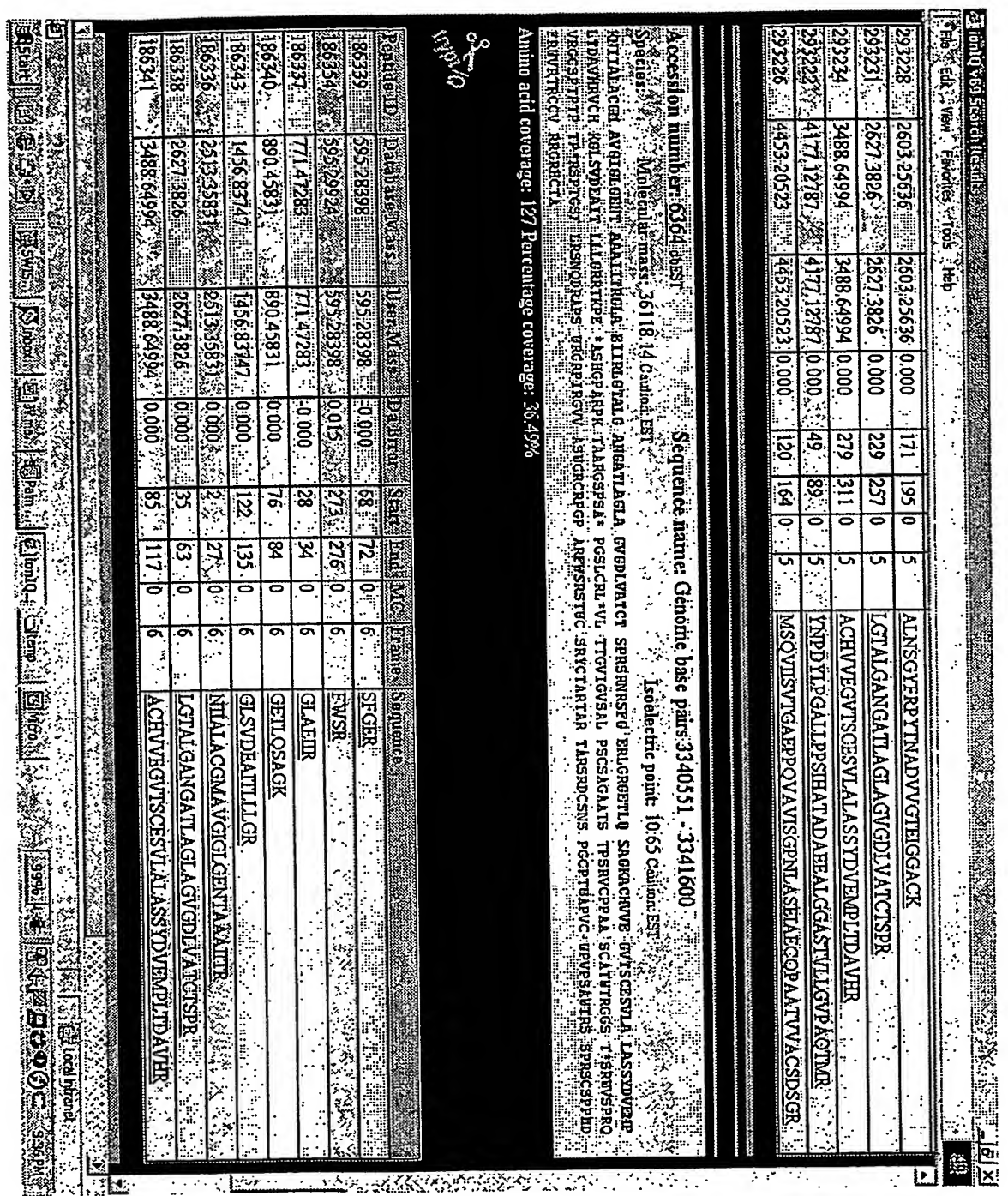
0-7-10
11/11/10

Episode ID	Datebase Mass	User Mass	Deletion	Start	End	MC	Frame	Sequence
186902	602.32618	602.32618	0.000	246	250	0	5	ANIER
186906	657.40476	657.40476	-0.000	321	326	0	5	AVAIOR
186904	989.54535	989.54535	0.000	267	275	0	5	GIELGIMR
186900	1217.61258	1217.61258	0.000	194	204	0	5	AEVADQINTIR
186899	1457.7752	1457.7752	0.000	179	192	0	5	VLADAGEVILMAR
186903	1665.95791	1665.95791	-0.000	251	266	0	5	WAPILPEGATVSLAR
186901	4177.12787	4177.12787	0.000	205	245	0	5	YNPDYLPGALLPRSHATADAEFATGGASTVILGVPAQIMR
186905	4453.20523	4453.20523	0.000	276	320	0	5	MSQVHSTYGAEPPOVAVISGPNILASEIACOPALATVACSDSGR

2005-25636-23 771-4728 FAX 978-595-28398 (5) 1505113488 04990 1800333740

Figure 5

Figure 6



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